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(54) Title: PROCESS FOR THE ISOLATION AND PURIFICATION OF MEVINOLIN**(57) Abstract**

The present invention relates to a process for the isolation of mevinolin by dissolving the active ingredient from the biomass into the fermentation liquor and subsequently separating it from the filtered fermentation liquor, which comprises carrying out the dissolution at a pH value between 7.5 and 10.0, preferably between 8.0 and 9.0, separating the active ingredient from the filtered liquor at a pH value between 4.5 and 1.0, preferably between 2.2 and 2.0, filtering and purifying it by methods known *per se*, preferably by recrystallization.

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PROCESS FOR THE ISOLATION AND PURIFICATION OF MEVINOLIN

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This invention relates to a process for the isolation and purification of mevinolin from fermentation liquor.

Mevinolin (lovastatin, monacolin K, MK 803) is a known antihypercholesterolemic agent, which can be produced by 10 fermentation using either a microorganism belonging to the species *Aspergillus terreus* or different microorganisms identified as species belonging to the *Monascus* genus.

The isolation of the active ingredient is carried out either by extracting directly the fermentation liquor with 15 a solvent or by extracting the filtered liquor and the biomass and subsequently purifying the crude product by chromatography.

For the extraction ethyl acetate, chloroform or benzene 20 is used. The fermentation liquor contains partly the open-chain hydroxy acid form of mevinolin, that is 3,5-dihydroxy-7-[1,2,6,7,8,8a-hexahydro-2,6-dimethyl-8-
-(2-methylbutyryloxy)-naphthalene-1-yl]-heptanoic acid.

This compound is heated in toluene to be lactonized to 25 mevinolin. The purification of the crude product containing mevinolin exclusively in the form of lactone is carried out by chromatography and subsequent recrystallization (US patent specification No. 4,319,039, Hungarian patent specifications No. 182,069, 182,075 and 187,296).

According to US patent specifications Nos. 4,231,938 30 and 4,319,039 beside the extraction an XAD₂ adsorption resin is also used for the isolation of mevinolin.

The main disadvantage of the extraction method resides 35 in the fact that the solvent dissolves, together with the active ingredient, a lot of concomitant contaminations rendering thereby the further purification more complicated and expensive. The purification at a proper efficiency can be accomplished namely by a multistage

column chromatographic method and subsequent re-crystallization.

Experiments have been carried out in order to compare the extraction method specified in Hungarian patent specification No. 187,296 to the method according to this invention for the isolation of mevinolin from fermentation liquor obtained by cultivation of an *Aspergillus obscurus* MV-1 holotype strain (deposition number: NCAIM (P)F 001189). The results of Example 1 prove that the product obtained from the fermentation liquor by extraction cannot be properly purified by recrystallization. The preparation of a product suitable for pharmaceutical purposes requires further purification by column chromatographic methods.

The present invention aims at providing a process for the isolation of mevinolin from fermentation liquor which can be carried out more readily and more economically than the hitherto known processes and enables the preparation of the active ingredient in a quality suitable for pharmaceutical purposes.

The present invention is based on the recognition that the active ingredient can be separated at high efficiency directly from the filtrate of the fermentation liquor (hereinafter: filtered liquor) at a pH value between 4.5 and 1.0. The crude product separated in this manner does not require to be purified by chromatography, as only a surprisingly slight amount of contamination separates together with it. Thus a simple recrystallization is sufficient to obtain a product of suitable quality.

According to the process of the invention the active ingredient is dissolved from the biomass into the fermentation liquor at a pH value between 7.5 and 10.0, the biomass is filtered off, the crude product is separated from the filtered liquor at a pH value between 4.5 and 1.0 and purified by methods known per se, preferably by recrystallization.

The separation of the active ingredient has been investigated at different acidic pH values. The pH range of 2.4 to 1.8, especially 2.2 to 2.0 has been found to be

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the most preferable. Besides, it has been found that the separation of the active ingredient from the filtered liquor, and especially the filterability of the precipitate can be improved by the addition of bivalent or trivalent metal salts, such as alkaline earth metal salts (CaCl_2 , MgCl_2 , MgSO_4) or earth metal salts [$(\text{Al}_2(\text{SO}_4)_3$].

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In order to support what has been said in the following Table data are given to show the active ingredient content of the filtered liquor after filtering off the active ingredient at different pH values in the presence of or without adding calcium chloride to the filtered liquor. The content of the active ingredient was determined by HPLC.

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pH	Active ingredient content of the filtered liquor (g/cm ³)	
	without adding any salt	in the presence of 0.2 M CaCl_2
7,0	418	60
6,0	387	65,8
5,0	201	103
4,0	58	50
3,0	31	22
2,0	14	10
1,5	10	10
1,0	8	8

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Taking into consideration that the majority of the active ingredient is bound to the biomass, both the efficiency of the dissolution into the fermentation liquor and the amount of the concomitant contaminations are of great importance.

Besides, it has also been recognized that by carrying out the dissolution of the active ingredient into the

fermentation liquor at a pH value between 7.5 and 10.0, particularly between 8.0 and 9.0 both the loss of substance and the amount of the concomitant contaminations can be reduced to a minimum.

5 According to our experiences the dissolution of the active ingredient can be enhanced by adding a slight amount of additives to the mixture. Aliphatic alcohols having 1 to 4 carbon atom(s), glycols having 2 to 5 carbon atoms, secondary or tertiary amines having 1 to 3 carbon atom(s), alkyl acetates having 1 to 5 carbon atom(s), dimethyl-formamide, polyethylene glycol or polypropylene glycol may serve as additives.

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In the following Table the active ingredient content of the filtered liquor is shown before the separation of the active ingredient at pH 9.0 and after the filtration thereof at pH 2.0 both in the presence of and without adding additives.

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Additive	Active ingredient content of the filtered liquor ($\mu\text{g}/\text{cm}^3$)	
1 % by vol.	pH: 9.0	pH: 2.0

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Diethylamine	412	9.2
Triethylamine	423	10.5
Dimethylformamide	460	6.9
Methanol	429	7.9
Ethanol	455	11.2
Isopropanol	467	8.7
Ethylene glycol	467	5.1
Propylene glycol	450	10.2
Polypropylene glycol	369	19.1
Isobutyl acetate	258	8.8
Polyethylene glycol	431	11.8
ontrol (without additive)	193	8.6

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From the data of the above Table it can be established that upon the addition of different additives the active ingredient content of the filtered liquor is higher than without using such additives. So the additives promote the dissolution of the active ingredient from the biomass into the fermentation liquor. At the same time it can also be seen that the additives do not have an influence on the separation, this latter can be performed at the same efficiency either in the presence of or without adding additives. The addition thereof is optionally reasonable, as they render the technological procedure simpler. In the presence of additives namely a single formation of a suspension from the biomass is sufficient, while without using additives this procedure has to be repeated in order to achieve the same efficiency.

For the purpose of additive ethylene glycol and ethanol are particularly preferred.

According to our experiences the additives effect their favourable activity even when applied in as slight amount as 0.1 % by volume calculated upon the volume of the fermentation liquor, and even when applied in greater amounts they do not have an influence on the separation of the dissolved active ingredient.

25	Concentration of ethanol % by volume	Active ingredient content of the filtered liquor ($\mu\text{g}/\text{cm}^3$)	
		pH: 9.0	pH: 2.0
30	0.1	400	8.9
	0.5	425	8.5
	1.0	455	11.2
	5.0	447	11.5
35	10.0	441	13.0
	15.0	434	18.1
	20.0	430	26.0

40 The crude product can be purified by any known method,

e.g. by a simple recrystallization. According to our experiments it is preferable to carry out the recrystallization from isobutyl acetate in such a manner that the solution of the substance in isobutyl acetate is washed with a weakly basic 2.5 w% ammonium sulfate solution adjusted to pH 8.5, the solvent phase is clarified with carbon, concentrated and the separated product is filtered off.

The advantages of the process according to the present invention are as follows: it renders possible the elimination of the extraction of both the fermentation liquor and the biomass from the technological procedure, the active ingredient separated from the filtered liquor at an acidic pH value is surprisingly pure, so it does not require to be purified by chromatography, but a simple recrystallization results in a product suitable for pharmaceutical purposes. Consequently the technological procedure is simple and can be accomplished economically, with a slight loss of substance (with a yield of higher than 90 %).

The process according to the invention can be applied by starting from any aqueous fermentation liquor cultured by a microorganism bio-synthesizing mevinolin either as the open-chain hydroxy acid or as lactone.

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The invention is illustrated in detail by the following Examples of non-limiting character:

Example 1

Comparative experiment according to the extraction method specified in Hungarian patent specification No. 187,296

800 g of fermentation liquor cultured by an Aspergillus obscurus MV-1 holotype strain (deposition number: NCAIM (P)F 001189) containing a total amount of 670 mg of mevinolin both as lactone and as hydroxy acid were adjusted to pH 4 with 20 wt% sulfuric acid solution. The liquor was then extracted with 400 cm³ of ethyl

acetate. The organic phase containing the active ingredient was separated and the aqueous residue was extracted again with further 400 cm³ of ethyl acetate. The ethyl acetate extracts were combined (760 cm³, active ingredient content: 643 mg), dried over anhydrous sodium sulfate and concentrated in vacuo. The concentrate was boiled in 100 cm³ of toluene for 2 hours. Then the undissolved particles were filtered off and washed successively with 50 cm³ of 5 wt% sodium hydrogen carbonate solution and

10 50 cm³ of water. The toluene solution was dried over anhydrous sodium sulfate and evaporated in vacuo. The active ingredient content of the thus-obtained 3.5 g of oily product amounted to 630 mg. In order to 15 crystallization the oily product was dissolved by warming in 15 cm³ of ethanol and allowed to stand at a temperature of 5°C for 24 hours. The product did not separate in crystalline form. The solvent was then removed and the oily product (3.5 g) was devideed into two parts.

20 1.75 g of product was recrystallized from 6 cm³ of isobutyl acetate as specified in Example 2. The product did not separate in crystalline form.

The other portion of the product was subjected to column chromatography using a column filled with 20 g of Kieselgel 60 (0.063 to 0.2 mm) (height: 22 cm, diameter: 1.6 cm). The column was eluted with a 40:60 mixture of ethyl acetate and methylene chloride at a rate of 20 cm³/hour. The 6 to 10 fractions containing the active ingredient were combined, clarified with activated carbon, 25 filtered and evaporated in vacuo to yield 260 mg of yellowish white solid residue, which was recrystallized from ethanol. The separated crystals were filtered through a G-4 sieve, washed with 10 cm³ of n-hexane and dried in vacuo at room temperature. Thus 180 mg of 30 chromatographically pure mevinolin were obtained. The evaporation residue of the mother liquor obtained during the recrystallization was recrystallized again from ethanol to obtain further 35 mg of mevinolin. The quality 35

of the product was the same as that of the first generation.

Example 2

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800 g of fermentation liquor cultured by an Aspergillus strain specified in Example 1 containing a total amount of 536 mg of mevinolin both as lactone and as hydroxy acid were diluted to 1200 g with water. Then the solution was
10 kept at a pH value between 8.5 and 9.0 with 20 wt% potassium hydroxide solution under continuous stirring for 2 hours. The biomass was then filtered off and suspended twice in 400 cm³ each of water. The suspension was adjusted to a pH value between 8.5 and 9.0 with 20 wt% potassium hydroxide solution, filtered again and the
15 filtrates were combined. Thus 1900 cm³ of filtered liquor containing 530 mg of active ingredient were obtained. The liquor was then adjusted to pH 2.1 with 15 wt% sulfuric acid solution, under stirring. The separated precipitate was settled, filtered, suspended in 100 cm³ of a sulfuric acid solution adjusted to pH 2 and filtered again. The active ingredient concentration of the filtrate amounted to 12 µg/cm³.

20 The filtered aqueous precipitate was dissolved in 50 cm³ of isobutyl acetate, the aqueous phase was separated and the solvent phase was concentrated to 2.5 cm³. The concentrate was dissolved in 60 cm³ of isobutyl acetate, washed twice with 60 cm³ each of an aqueous ammonium sulfate solution adjusted to pH 8.5 with ammonium
25 hydroxide, clarified with 0.5 g of carbon, concentrated to 10 cm³, allowed to crystallize for 24 hours at 5°C, filtered and dried in vacuo. Thus 436 mg of mevinolin were isolated. Active ingredient content: 98.7 % (HPLC).

30 From the combined mother liquors further 65 mg of mevinolin were obtained in a purity of 92.8 %. The crude products were combined and recrystallized from ethanol. Thus 450 mg of product were isolated.
35 Active ingredient content: 99.8 % (HPLC).

Dihydromevinolin content: 0.17% (GC)
[α] $25_D = +329.8^\circ$ (c=0.5; acetonitrile)

Example 3

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800 g of fermentation liquor cultured by an Aspergillus strain specified in Example 1 containing a total amount of 605 mg of mevinolin both as lactone and as hydroxy acid were diluted to 1200 g with water. Then 2,4 g of ethylene glycol were added to the mixture, and it was kept at a pH value between 8.5 and 9.0 by adding 20 wt% potassium hydroxide solution under continuous stirring for 2 hours. The biomass was then filtered off and suspended in 400 cm³ of water containing 0.8 g of ethylene glycol. The suspension was adjusted to a pH value between 8.5 and 9.0 with 20 wt% potassium hydroxide solution, filtered again and the filtrates were combined. The thus-obtained 1470 cm³ of filtered liquor containing 600 mg of active ingredient were adjusted to pH 2.1 with 15 wt% phosphoric acid under stirring. The precipitate was settled for 4 hours. Further on the process specified in Example 2 was followed.

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Thus 548 mg of mevinolin were isolated.

Active ingredient content: 99.7 % (HPLC).

Dihydromevinolin content: 0.15 % (GC)

[α] $25_D = +329^\circ$ (c=0.5; acetonitrile)

Example 4

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800 g of fermentation liquor cultured by an Aspergillus strain specified in Example 1 containing a total amount of 575 mg of mevinolin both as lactone and as hydroxy acid were diluted to 1200 g with water. Then 2,4 g of ethylene glycol were added to the mixture, and the pH were kept at 9.0 to 9.5 by adding 20 wt% potassium hydroxide solution under continuous stirring for 2 hours. The biomass was then filtered off and suspended in 400 cm³ of water. The suspension was adjusted to a pH value between 9.0 and 9.5

with 20 wt% potassium hydroxide solution, filtered again and the filtrates were combined. Thus 1480 cm³ of filtered liquor containing 567 mg of active ingredient were obtained. Then 3.5 g of calcium chloride were added to it and the solution was adjusted to pH 2.1 with 15 wt% sulfuric acid solution under stirring. The separated precipitate was settled for 4 hours. Further on the process specified in Example 2 was followed, with the difference that the active ingredient was dissolved from the precipitate with 120 cm³ of isobutyl acetate.

Thus 527 mg of mevinolin were isolated.

Active ingredient content: 99.2 % (HPLC).

Dihydromevinolin content: 0.25% (GC)

[α] 25_D = +329.5° (c=0.5; acetonitrile)

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Example 5

10000 g of fermentation liquor cultured by an Aspergillus strain specified in Example 1 containing a total amount of 4180 mg of mevinolin both as lactone and as hydroxy acid were diluted to 15000 g with water. Then 30 g of ethylene glycol were added to the mixture, and it was kept at a pH value between 8.0 and 8.5 by adding 20 wt% potassium hydroxide solution under continuous stirring for 2 hours. The biomass was then filtered off and suspended in 5 dm³ of water containing 10 g of ethylene glycol. The suspension was adjusted to a pH value between 8.0 and 8.5 with 20 wt% potassium hydroxide solution, filtered again and the filtrates were combined. Thus 18200 cm³ of filtered liquor containing 4091 mg of active ingredient were obtained. Then 20 g of magnesium sulfate were added to the mixture and it was adjusted to pH 2.1 with 15 wt% sulfuric acid solution, under stirring. The separated precipitate was settled, filtered, suspended in 1200 cm³ of an aqueous sulfuric acid solution adjusted to pH 2 and filtered again. The filtered aqueous precipitate was dissolved in 600 cm³ of isobutyl acetate, the aqueous phase was separated and the solvent phase was concentrated

to 30 cm³. The concentrate was dissolved in 400 cm³ of isobutyl acetate, washed twice with 400 cm³ each of 2.5 wt% ammonium sulfate solution adjusted to pH 8.5 with ammonium hydroxide solution and clarified with 6 g of carbon by stirring for half an hour at room temperature. The solution was concentrated to 80 cm³, allowed to crystallize for 24 hours at 5°C, filtered and dried in vacuo. Further on the process specified in Example 2 was followed.

Thus 3432 mg of mevinolin were isolated.

Active ingredient content: 99.1% (HPLC).

Dihydromevinolin content: 0.19 % (GC)

[α] 25D = +328.9° (c=0.5; acetonitrile)

15 Example 6

100 kg of fermentation liquor cultured by an *Aspergillus* strain specified in Example 1 containing a total amount of 44.3 g of mevinolin both as lactone and as hydroxy acid were diluted to 150 kg with water. Then 300 g of ethylene glycol were added to the mixture, and it was kept at a pH value between 8.5 and 9.0 by adding 20 wt% potassium hydroxide solution under continuous stirring for 2 hours. The biomass was then filtered off and suspended in 50 kg of water containing 100 g of ethylene glycol. The suspension was adjusted to a pH value between 8.5 and 9.0 with a 20 wt% potassium hydroxide solution, filtered again and the filtrates were combined. Thus 183 kg of filtered liquor containing 42.9 g of active ingredient were obtained. Then 200 g of magnesium sulfate were added to it and the solution was adjusted to pH 2.1 with 15 wt% sulfuric acid solution, under stirring. The separated precipitate was settled filtered, suspended in 12 dm³ of a sulfuric acid solution adjusted to pH 2 and filtered again. The filtered aqueous precipitate was dissolved in 6 dm³ of isobutyl acetate, the aqueous phase was separated and the solvent phase was concentrated to 300 cm³. The concentrate was dissolved in 4 dm³ of isobutyl acetate,

washed twice with 4 dm³ each of 2.5 wt% ammonium sulfate solution adjusted to pH 8.5 with ammonium hydroxide solution and clarified with 60 g of carbon by stirring for half an hour at room temperature. The solution was concentrated to 0.8 dm³, allowed to crystallize for 24 hours at 5°C, filtered and dried in vacuo. Further on the process according to Example 2 was followed.

5 Thus 37.03 g of mevinolin were isolated.

Active ingredient content: 99.3% (HPLC).

10 Dihydromevinolin content: 0.18 % (GC)

[α] 25_D = +329.5° (c=0.5; acetonitrile)

What we claim is:

1. A process for the isolation of mevinolin by dissolving the active ingredient from the biomass into the fermentation liquor and subsequently separating it from the filtered fermentation liquor, which comprises carrying out the dissolution at a pH value between 7.5 and 10.0, preferably between 8.0 and 9.0, separating the active ingredient from the filtered liquor at a pH value between 4.5 and 1.0, preferably between 2.2 and 2.0, filtering and purifying it by methods known per se, preferably by recrystallization.
2. A process as claimed in claim 1, which comprises carrying out the dissolution in the presence of any of the following additive(s) applied in an amount of at least 0.1 wt% related to the volume of the fermentation liquor: aliphatic alcohols having 1 to 4 carbon atom(s), glycols having 2 to 5 carbon atoms, secondary or tertiary amines having 1 to 3 carbon atom(s), alkyl acetates having 1 to 5 carbon atom(s), dimethylformamide and/or polyethylene glycol and/or polypropylene glycol.
3. A process as claimed in claim 2, which comprises using as additive ethanol or ethylene glycol.
4. A process as claimed in any of claims 1 to 3, which comprises adding an alkaline earth metal salt or an earth metal salt to the filtered liquor prior to the separation.

INTERNATIONAL SEARCH REPORT

International application No.

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A. CLASSIFICATION OF SUBJECT MATTER

IPC⁵: C 12 P 17/06

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C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category ^a	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim N .
A	DE, A1, 3 006 215 (SANKYO) 27 November 1980 (27.11.80), example 1.	1
A	DE, A1, 3 006 216 (SANKYO) 04 September 1980 (04.09.80), example.	1
A	Proceedings of the National Academy of Sciences of the United States of America, Volumne 77, no. 7., published July 1980 (Baltimore, USA), A.W. ALBERTS et al. "Mevindin: A highly potent competitive inhibitor of hydroxymethylglutaryl-coenzyme A reductase and a cholesterol-lowering agent", pages 3957-3961, especially pages 3957, 3958. -----	1

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Date of the actual completion of the international search 17 December 1993 (17.12.93)	Date of mailing of the international search report 03 January 1994 (03.01.94)
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NO A	800451	21-08-80
NO B	153974	17-03-86
NO C	153974	25-06-86
NZ A	192919	06-07-84
PH A	15145	24-08-82
PL A1	222120	20-10-80
PL B1	124304	31-01-83
SE A	8001339	21-08-80
SE B	453301	25-01-88
SE C	453301	06-04-89
SG A	67/84	08-02-85
SU A1	969702	30-10-82
SU A3	1158048	23-05-85
ZA A	8000962	25-03-81
KR A	8302801	16-12-83